

学術情報リポジトリ

Materials for Pythium Flora of Japan (VI): Morphology of acquired resistant isolates of Pythium vanterpoolii against metalaxyl

メタデータ	言語: English
	出版者:
	公開日: 2009-08-25
	キーワード (Ja):
	キーワード (En):
	作成者: ICHITANI, Takio, FUJITA, Yuriko, KOBAYASHI,
	Toshimi
	メールアドレス:
	所属:
URL	https://doi.org/10.24729/00009215

Materials for Pythium Flora of Japan (VI) Morphology of acquired resistant isolates of *Pythium vanterpoolii*against metalaxyl

Takio ICHITANI, Yuriko FUJITA and Toshimi KOBAYASHI

Laboratory of Plant Pathology, College of Agriculture

(Received, 1993)

Abstract

There were no significant differences in morphology, with special reference to the size of oogonium and oospore, between isolate UOP 393 of *Pythium vanterpoolii* with acquired resistance to metalaxyl and its sensitive parent isolate UOP 392.

Pythium vanterpoolii is the most important pathogen causing Pythium blight of zoysia grass in golf course greens¹⁾. A field isolate UOP 392 of this fungus sensitive to metalaxyl was originally obtained from a basal part of an erect stem of diseased zoysia in the No. 12 green of Meisho Golf Club (Mie Prefecture, Japan) in April 29, 1989. This isolate, together with eight others, became resistant during 14-15 transfers on CMA medium amended with sublethal concentrations of metalaxyl²⁾ and was designated as an acquired resistant isolate UOP 393³⁾. Its resistance, cultural characteristics and virulence were retained for two years although most of the other isolates partially or completely lost their resistance⁴⁾. P. vanterpoolii UOP 393 could be maintained for a further year on CMA medium without any great change in these cultural characteristics⁵⁾, and proved to possess acquired resistance under glasshouse conditions⁵⁾.

This papaer deals with the detailed description of the oogonium and oospore of the acquired resistant isolate UOP 393, following the method reported previously^{6,7)} and in comparison with the sensitive parent isolate UOP 392.

Table 1. Comparison of mycelial growth rate and colony type between acquired resistant isolate UOP 393 and sensitive parent isolate UOP 392 of P. vanterpoolii grown on Bacto-PDA medium at 25°C.

Isolate no.	Exp.	Mye (Culture	Colony type		
		1	2	3	
	1	17.8	21.8	23.1	A
UOP 393	2	17.0	22.4	24.4	A
	Average	17.4	22.1	23.8	
	1	20.0	23.5	25.0	A
UOP 392	2	20.0	24.8	26.4	A
	Average	20.0	24.2	25.7	

Acquired resistant isolate of Pythium vanterpoolii UOP 393

General morphology The morphology of this isolate was essentially the same as that reported previously^{8,9)}. In particular, the catenulate sporangia (Fig. 1) were the most characteristic feature of this isolate, as V. & H. Kouyeas⁹⁾ have already pointed out for *P. vanterpoolii*.

Growth rate and colony morphology No significant differences were found in growth rate and colony morphology (Fig. 3) between the acquired resistant (UOP 393) and sensitive parent (UOP 392) isolates (Table 1).

Morphology of oogonium and oospore As shown in Table 2, there were no significant differences in size of oogonium and oospore, and thickness of oospore wall

Table 2. Comparison of size of oogonium and oospore, and thickness of oospore wall between acquired resistant isolate UOP 393 and sensitive parent isolate UOP 392 of *P. vanterpoolii* grown on Bacto-CMA medium at 25°C a).

Culture period (days)	Diam. (µm)	of oogonium	Diam. (µm)	of oospore	Thickness (µm) of oospore wal		
	UOP 393	UOP 392	UOP 393	UOP 392	UOP 393	UOP 392	
3	22.67 (1.62)	22.85 (1.65)		_		_	
5	22.67 (1.69)	22.59 (1.67)	_	_	_	_	
6	22.84 (1.59)	22.67 (1.55)	19.75 (1.02)	20.65 (0.80)	_	_	
8	_	_	20.92 (1.12)	20.87 (1.00)	1.86 (0.19)	1.79 (0.29)	
20		_	20.80 (1.26)	20.71 (1.02)	1.93 (0.19)	2.06 (0.18)	
30	-	_	20.92 (1.14)	20.86 (1.14)	2.06 (0.15)	2.08 (0.16)	
Average	22.73	22.70	20.60	20.77	1.95	1.98	

a) (): Standard deviation, -: not measured.

Table 3-1. Comparison of mycelial growth rate and colony type between hyphal tip cultures of acquired resistant isolate UOP 393 and sensitive parent isolate UOP 392 of *P. vanterpoolii* grown on Bacto-PDA medium at 25°C ^{a)}.

Hyphal tip culture no. of UOP 393	Myc (Culture	elial gro days; m		Colony Hyphal tip Mycelial growth type culture no. (Culture days; mm/24hr)	, , , , , , , , , , , , , , , , , , ,	, ,, , , , , , , , , , , , , , , , , ,	•		Colony r) type
	1	2	3		of UOP 392	1	2	3	
R-I	14.8	17.1	18.5	В	S-1	20.0	23.5	24.7	Α
R-2	18.0	22.0	23.5	Α	S-2	19.8	23.9	24.9	Α
R-3	13.8	15.6	16.1	В	S-3	19.5	23.3	24.7	Α
R-4	13.0	16.1	17.8	В -	S-4	19.8	23.5	24.8	Α
R-5	18.8	22.4	23.8	Α	S-5	19.5	23.6	24.8	Α
R-6	14.8	16.9	18.1	В	S-6	19.5	23.8	24.5	Α
R-7	19.0	22.5	23.8	Α	S-7	19.5	23.4	24.6	Α
R-8	17.0	20.0	20.9	Α	S-8	19.0	23.0	24.1	Α
R-9	13.5	15.8	16.7	В	S-9	20.0	23.8	24.8	Α
R-10	18.0	21.6	22.9	Α	S-10	20.0	23.9	25.0	Α
R-11	18.3	22.3	23.3	Α	S-11	20.2	24.1	25.2	Α
Average	16.3 (2.20)	19.3 (2.84)	20.5 (2.94)		Average	19.7 (0.33)	23.6 (0.31)	24.7 (0.27)	

a) (): Standard deviation

Table 3-2. Comparison of mycelial growth rate and colony type between hyphal tip cultures of acquired resistant isolate UOP 393 and sensitive parent isolate UOP 392 of *P. vanterpoolii* grown on Bacto-PDA medium at 25°C a).

Hyphal tip culture no. of UOP 393		•	Mycelial growth culture days; mm/24hr)		Colony type			Mycelial growth (Culture days; mm/24hr)		
	1	2	3		of UOP 392	1	2	3		
R-21	14.0	17.0	19.2	В	S-21	19.0	24.5	26.2	Α	
R-22	21.3	26.0	28.0	Α	S-22	20.7	26.2	27.4	Α	
R-23	17.8	23.1	25.2	$_{0}\mathbf{A}$	S-23	18.0	24.3	26.3	Α	
R-24	18.8	23.9	24.9	Α	S-24	21.8	26.3	27.5	Α	
R-25	12.0	17.4	19.5	В	S-25	20.5	25.6	27.1	Α	
R-26	16.0	20.3	22.6	В	S-26	20.5	26.8	28.7	Α	
R-27	18.5	23.5	25.2	Α	S-27	17.5	23.3	25.7	Α	
R-28	16.5	20.8	22.3	В	S-28	18.5	23.6	25.1	Α	
R-29	19.3	23.6	26.1	Α	S-29	17.8	23.8	25.8	Α	
Average	17.1 (2.69)	21.7 (2.90)	23.7 (2.81)		Average	19.4 (1.45)	24.9 (1.23)	26.6 (1.06)		

a) (): Standard deviation

Table 4-1. Comparison of size of oogonium and oospore, and thickness of oospore wall between hyphal tip cultures (R-22, R-26, R-28) of acquired resistant isolate UOP 393 of *P. vanterpoolii* grown on Bacto-CMA medium at 25°C a).

Culture period (days)	Diam. (um) of o	ogonium	Diam.	Diam. (μm) of oospore			Thickness (µm) of oospore wa			
	R-22	R-26	R-28	R-22	R-26	R-28	R-22	R-26	R-28		
3	22.28 (1.73)	23.69 (1.77)	23.83 (1.58)	_	_	_			_		
5	22.58 (1.56)	23.88 (1.84)	23.83 (1.64)			_	_	_	_		
6	22.58 (1.62)	23.63 (1.77)	23.39 (1.77)	21.07 (1.47)		_	1.69 (0.25)	_	_		
8	· -	23.40 (1.81)	23.83 (1.70)	20.79 (1.37)	21.61 (1.34)	21.61 (1.36)	1.81 (0.16)	1.58 (0.23)	1.62 (0.22)		
20	_	_	_	20.64 (1.44)	21.51 (1.22)	21.40 (1.34)	1.93 (0.12)	1.82 (0.15)	1.79 (0.14)		
30	_		_	20.40 (1.56)	21.46 (1.23)	21.49 (1.20)	1.98 (0.10)	1.92 (0.14)	1.92 (0.13)		
Average	22.48	23.65	23.72	20.73	21.53	21.50	1.85	1.77	1.78		

a) (): Standard deviation, -: not measured.

Hyphal tip cultures from acquired resistant isolate UOP 393 of *P. vanterpoolii*General morphology The morphology of the hyphal tip cultures was same as that of the parent acquired resistant isolate. The catenulate sporangia (Fig. 1) were the most characteristic feature.

Growth rate and colony morphology No significant differences were found in growth rate among hyphal tip cultures of acquired resistant isolate UOP 393 and sensitive parent isolate UOP 392. Similarly, no remarkable differences were found in growth rate of hyphal tip cultures between acquired resistant isolate UOP 393 and sensitive parent

⁽Fig. 2) between two isolates.

Table 4-2.	Comparison of size of oogonium and oospore, and thickness of oospore wall between hyphal
	tip culture (R-22) of acquired resistant isolate UOP 393 and those (S-22, S-28) of sensitive
	parent isolate UOP 392 of P. vanterpoolii grown on Bacto-CMA medium at 25°C a).

Culture period (days)	Diam. (µm) of oogonium			Diam.	Diam. (μm) of oospore			Thickness (µm) of oospore wall			
	R-22	S-22	S-28	R-22	S-22	S-28	R-22	S-22	S-28		
3	22.93 (1.56)	23.04 (1.67)	22.95 (1.67)	_		_	_		-		
5	22.99 (1.52)	23.13 (1.62)	23.06 (1.51)		. –		- ,	-	_		
6	22.83 (1.59)	22.95 (1.53)	22.85 (1.67)	21.09 (1.41)	21.12 (1.42)	20.93 (1.49)	1.62 (0.30)	1.65 (0.29)	1.60 (0.28)		
8			_	20.96 (1.48)	21.14 (1.43)	21.10 (1.42)	1.78 (0.19)	1.72 (0.23)	1.73 (0.19)		
20	_ '		_	20.44 (1.29)	20.90 (1.56)	21.12 (1.44)	1.92 (0.13)	1.91 (0.14)	1.94 (0.13)		
30	_	_	_	20.24 (1.42)	20.65 (1.83)	20.88 (1.38)	1.97 (0.11)	1.96 (0.12)	1.97 (0.12)		
Average	22.92	23.04	22.95	20.68	20.95	21.01	1.82	1.81	1.81		

a) (): Standard deviation, -: not measured.

isolate 392 (Tables 3-1, 2). Most of their colonies were arachnoid in habit with profuse aerial mycelium, some with moderate aerial mycelium (Fig. 3).

Morphology of oogonium and oospore As shown in Tables 4-1, 2, there were no significant differences in size of oogonium and oospore, and thickness of oospore wall among hyphal tip cultures of acquired resistant isolate UOP 393 and sensitive parent isolate UOP 392. Similarly, no significant differences were found in morphology of hyphal tip cultures between acquired resistant isolate UOP 393 and sensitive parent isolate UOP 392.

From the results obtained, there were no significant differences in morphology, with special reference to the size of oogonium and oospore, between isolate UOP 393 of *P. vanterpoolii* with acquired resistance to metalaxyl and the sensitive isolate UOP 392.

The authors are indebted to Dr. D. J. Stamps, formerly of the CAB International Mycological Institute, Kew, UK, for critical reading of the manuscript.

References

- 1) ICHITANI, T., TANI, T. and UMAKOSHI, T. (1986). Identification of *Pythium* spp. pathogenic on manila grass (*Zoysia matrella* Merr.). *Trans. mycol. Soc. Japan*, 27, 41-50 (in Japanese with English summary).
- 2) ICHITANI, T., FUJII, K., TANPO, H. and TANI, T. (1991). Resistance to metalaxyl in *Pythium vanterpoolii* causing two different turf diseases of zoysia greens. *Journ. Japanese Soc. Turfgrass Sci.*, 20, 33-40 (in Japanese with English summary).
- 3) FUJITA, Y., ICHITANI, T, and TANI, T. (1993). Cultural characteristics of acquired

- resistant isolate of Pythium vanterpoolii against metalaxyl. Ann. Phytopath. Soc. Japan, 59, 77 (Abstract in Japanese).
- 4) ICHITANI, T., FUJITA, Y., HAMANAKA, M. and TANI, T. (1993). Differences in cultural characteristics, virulence and loss of resistance of acquired resistant isolate of *Pythium vanterpoolii* against metalaxyl. *Journ. Japanese Soc. Turfgrass Sci.*, 21, 242 (Abstract in Japanese).
- 5) FUJITA, Y., ICHITANI, T. and TANI, T. (1993). Cultural characteristics and virulence of acquired resistant isolate of *Pythium vanterpoolii* against metalaxyl. *Journ. Japanese Soc. Turfgrass Sci.*, 22, 13-20 (in Japanese with English summary).
- 6) ICHITANI, T. and KANG, H. T. (1988). Materials for Pythium Flora of Japan (I) Plant-pathogenic Pythium irregulare. Bull. Univ. Osaka Pref., 40, 19-26.
- 7) ICHITANI, T., TAKAMATSU, S. and STAMPS, D. J. (1986). Identification and pathogenicity of three species of *Pythium* newly isolated from diseased wheat and barley just after thawing in Japan. *Ann. Phytopath. Soc. Japan*, 52, 209-216.
- 8) ICHITANI, T., KANG, H. T. and MINE, K. (1989). Materials for Pythium Flora of Japan (II) *Pythium torulosum* and *P. vanterpoolii* from golfgreens of manilagrass or bentgrass. *Bull. Univ. Osaka Pref.*, 41, 9-19.
- 9) KOUYEAS, V. and KOUYEAS, H. (1963). Notes on species of *Pythium. Ann. Inst.* phytopath. Benaki, N. S., 5, 207-237.

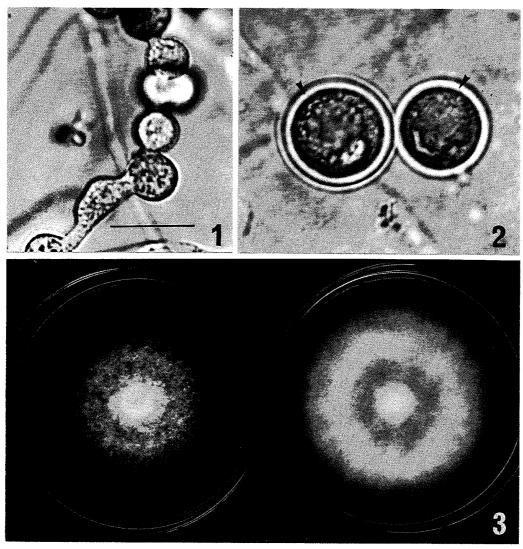


Fig. 1 Catenulate sporangia.

Fig. 2 Plerotic oospores, with relatively thick wall (arrows).

Fig. 3 Colony types A (right) with arachnoid habit and profuse aerial mycelium, and B (left), the same habit and moderate aerial mycelium. Bar (20 μ m) on fig. 1 is also applicable to fig. 2.